Review

The Importance of Proteins in Defense Against Oxidation

EMMANUEL BOURDON and DENIS BLACHE

ABSTRACT

Free radicals are a normal feature of cellular oxygen metabolism. However, free radical-associated damage is an important factor in many pathological and toxicological processes. For a long time, lipid peroxidation, mediated by oxygen-derived free radicals, was probably the most extensively investigated process. From more recent studies, it has become evident that proteins are also the targets of free radicals, and this has important implication for their activity, unfolding, and degradation, as well as in cell functioning. After giving a brief overview of the key role of proteins in the overall antioxidant defense, this review examines their role as targets of oxidation reactions, taking into account the reactivity of amino acid residues and some of their oxidation products. In light of recent data, we then consider the specific role of sulfur-containing amino acids in protein degradation and their possible interplay with the reversal of limited oxidative lesions. The participation of proteins in the overall antioxidant defense is also discussed, specifically the role of metallothionein as an intracellular antioxidant and that of albumin as a circulating antioxidant. Antioxid. Redox Signal. 3, 293–311.

INTRODUCTION

 ${f R}^{{\scriptsize {\sf EACTIVE}}}$ OXYGEN SPECIES (ROS) is a collective term for oxygen radicals and nonradicals that are oxidizing agents and/or are easily converted into free radicals. Because ROS can be generated in vivo, recent advances in our understanding of the biochemistry of these compounds have led to the recognition that ROS are likely implicated in the pathogenesis of a number of diseases, including cancer and cardiovascular and neurodegenerative diseases, as well as in certain physiological processes such as aging. Oxidative stress occurs when endogenous antioxidant defenses are inadequate to scavenge ROS completely. The resulting oxidative damage to lipids, DNA, proteins, and other molecules may contribute to the development of the aforementioned diseases (58). It follows that there is a growing interest in additional research on biomarkers (31) and mechanisms of antioxidant protection with potential relevance for therapeutic use (32, 57).

In this review, we discuss the role of proteins as targets of free radical attack, as well as their role as part of the antioxidant defense in plasma and various intracellular compartments. With a special focus on the oxidation ability of amino acids, we shall highlight the importance of sulfur-containing amino acid residues. As examples, the specific roles of serum albumin and metallothionein (MT) will be especially described.

THE ANTIOXIDANT DEFENSE

ROS are produced either endogenously, from normal metabolic reactions, or exogenously as components of tobacco smoke and air

pollutants (21, 88, 100) as well as indirectly through the metabolism of drugs, solvents, pesticides, and exposure to radiation (UV light) (5, 57–59, 76). The cytochrome *c* oxidase-catalyzed reaction involves the transfer of electrons to oxygen and results in partially reduced oxygen species. Other enzymes, especially flavin enzymes, also generate partially reduced oxygen species. However, numerous natural defenses exist either to prevent ROS formation or to neutralize them after they are generated (26, 57). They include enzymes and other low-molecular-weight compounds. The discovery of superoxide dismutase (SOD) provided understanding of the importance of free radicals in biological systems (49, 60). Three distinct forms have been described: CuZn-SOD, which is cytosolic; Mn-SOD, which is located in mitochondria; and a third form, also a CuZn-SOD, which is immunologically distinct from the others and is located extracellularly. SOD catalyzes the conversion of superoxide anion $(O_2^{-\bullet})$ to hydrogen peroxide (H_2O_2) . In the presence of transition metals, H₂O₂ is rapidly converted to the potent HO' through the Fenton reaction. Fortunately, two enzymes are able to neutralize H₂O₂: catalase and glutathione peroxidase. The latter enzyme also has the properties to reduce peroxides, including lipid peroxides. It has been shown, however, that increased activity/expression of only one antioxidant enzyme could be deleterious and that it is the balance between CuZn-SOD and the peroxide-removing enzymes catalase and glutathione peroxidase that is of prime importance in the antioxidant defense. An additional antioxidant defense mechanism is to keep transition metal ions safely sequestered by metal binding proteins, such as ferritin, transferrin, ceruloplasmin, MT, and possibly albumin.

Aside from these enzymes and proteins, an array of antioxidant molecules are present to scavenge free radicals (2). The most notable are vitamins C and E, various carotenoids (β -carotene, lycopene), glutathione, ubiquinol, uric acid, and bilirubin. It is interesting to note that several dietary micronutrients contribute greatly to the protective mechanism. This is particularly true for vitamins, oligominerals and perhaps also phytochemicals such as polyphe-

nols (flavonoids, resveratrol), which are currently the subject of extensive research (3).

Another defense system that has been often neglected also exists (83). It consists of various enzymes involved in the repair of ROS-induced damage to biomolecules. It includes the following: (i) phospholipases (phospholipases A2, platelet-activating factor-acetyl hydrolase), which remove the oxidized products of fatty acids because generally the latter have to be in a nonesterified form to be detoxified (80, 110, 118, 123); (ii) nucleases, also named redoxyendonucleases (33), which repair lesions occurring in DNA either via endogenously generated oxidant or following exposure to ionizing radiations (83); and (iii) proteases generated from latent proteolytic systems (26, 95) activated when cells have been challenged with ROS for the removal of oxidatively modified proteins (see below).

ROLE OF PROTEINS IN THE ANTIOXIDANT ACTIVITY OF SERUM

Endogenous ROS include those that are produced intracellularly as well as those that are released into the surrounding area. Defenses against ROS are generally located close to the site of production. However, circulating fluids not only bring the nutrients necessary for cellular metabolism, they also help to neutralize cell-borne ROS and to improve cellular defenses. Previous work has shown that blood serum has strong antioxidant properties (124). This has been demonstrated by incubations of tissue homogenates or lipid emulsions in free radical-producing systems. Surprisingly, most of the serum's protecting activity has not been attributed to vitamin E or C, but to the presence of urate and proteins (17, 75, 124). Our laboratory has obtained similar results using 2,2'-azo-bis(2-amidinopropane) dihydrochloride (AAPH)-mediated blood hemolysis (8, 35, 51). We found that diluted serum strongly inhibited the free radical-induced hemolysis of washed red blood cells (Fig. 1). The contribution of various components of serum were analyzed using different experimental conditions as shown in Table 1. These data showed that

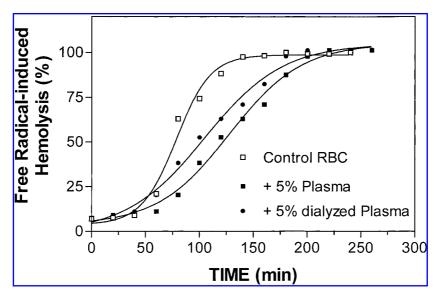


FIG. 1. Antioxidant effect of plasma. Washed red blood cells (RBC) were incubated in the presence of an azo-typed free radical generator as described in the text (34, 51). Plasma or dialyzed plasma was added in a final 5% volume, and the time course of the hemolysis (in %) was plotted and fitted by computer analysis. Curves were characterized by the time corresponding to 50% hemolysis and are from one representative experiment carried out with the same plasma in Table 1. For the purpose of clarity, standard deviations have not been represented. Results (n = 3) are 78.3 ± 2.8 , 127.7 ± 3.8 , and 104.8 ± 4.8 for control RBC, 5% plasma, and 5% dialyzed plasma, respectively.

20–25% of the total antioxidant activity of serum was due to urate, as assessed by either removal of urate (and other small molecules) by dialysis or addition of urate to levels found in lipoprotein-deficient serum. The contribution of vitamins E and C was obtained by the difference in antioxidant activity between total

serum and lipoprotein-deficient serum because these were removed during preparation of lipoprotein-deficient serum. Consequently, most of the total antioxidant activity of whole serum can be attributed to proteins, especially albumin. Similar results were obtained with dialyzed lipoprotein-deficient serum and equiv-

Table 1. Free Radical-Induced Hemolysis and Serum Antioxidants

	HT 50% (min)	%
Red blood cells (control)	74	100
+ 5% Serum	129	174
+ lipoprotein-deficient serum (dialyzed)	109	147
+ lipoprotein-deficient serum + ascorbic acid (2 μM)	112	151
+ lipoprotein-deficient serum + urate (15 μM)	130	175
+ HSA (2.5 g/L)	115	155
+ urate $(15 \mu M)$	96	130
+ HSA (2.5 g/L) + urate (15 μ M)	126	170

Washed human red blood cells were incubated with an azo derivative as free radical generator, and samples were analyzed for the released hemoglobin as in Fig. 1. Results are expressed as half hemolysis time (HT 50%) expressed in min. Various additions, such as 5% diluted serum, ascorbic acid, uric acid, and serum albumin (HSA), were performed and compared with control conditions (red blood cells alone). Antioxidants were added in concentrations equivalent to those found in 5% diluted serum. Data (average values, with standard deviations <7%) are representative of several experiments carried out with the same serum having the following characteristics: HSA, 48 g/L; ascorbic acid, 34 μ M; uric acid, 300 μ M. Dialyzed serum has no detectable ascorbic acid and uric acid.

alent solutions of human serum albumin against free radical-induced hemolysis (Table 1). These data confirm earlier findings on the measurement of individual antioxidant and radical trapping activity of plasma, using oxygen uptake (18, 19, 124). It was also concluded that the thiol content of plasma could account for 52–80% of the total radical trapping activity.

OXIDATIVE MODIFICATION OF AMINO ACIDS AND PROTEINS

A molecule that could intercept the oxidation reaction is regarded as an antioxidant if this interception interrupts the propagation reaction. Proteins may be considered as such because, although the prime targets for ROS are polyunsaturated fatty acids, proteins are also very sensitive to direct peroxidative damage. Proteins may work as sacrificial antioxidants. The consequences are that these free radical-mediated modifications may alter the metabolism and the function of the target proteins.

It has largely been shown that isolated amino acids can be oxidatively modified (Table 2). The quantitative significance of the overall oxidative process, involving either ionizing radiations or metal-catalyzed oxidations, is dependent upon the complexity of the amino acids examined (aliphatic, aromatic), the availability of oxygen, and the strength of the oxidation reaction (109). The resulting products are α -keto acids, hydroxyamino acids accompanied by a mixture of cross-linked derivatives. Formyl-kynurenine is a major product of tryptophan

oxidation, whereas *o*-tyrosine and other hydroxy derivatives are formed from phenylalanine. Tyrosine oxidation may also lead to dimerization and to formation of 3,4-dihydroxyphenylalanine (DOPA). Among the various amino acids, sulfur-containing ones such as methionine and cysteine are considered as preferential targets. Methionine is almost completely recovered as methionine sulfoxide or sulfone (Fig. 2) (122).

The oxidative modifications of amino acid residues may lead to important consequences for the protein concerned. It may vary from slight modification of the protein structure to large denaturation accompanied by fragmentation. The most relevant studies have established that histidine, arginine, lysine, proline, methionine, and cysteine residues are among the most common sites of oxidation by metalcatalyzed processes, whereas all amino acid residues are susceptible to oxidation by ROS generated during radiolysis. Because histidine preferentially works as a ligand for divalent metals, the resulting oxidation product, 2-oxohistidine, may be used as a marker for metalcatalyzed oxidative modifications occurring in the vicinity of the binding site of the protein (71). Figure 3 shows a proposed mechanism for the iron-catalyzed site-specific oxidation of the ϵ -amino group of a lysine residue in a protein (109). The immediate environment of proteins influences the nature and extent of their reactions with ROS. The radical-induced modifications depend on protein concentration and on radical flux and specificity (30). Lipid oxidation products such as aldehydes can react with

Table 2. Some Oxidation Products of Amino Acids

Amino acids	Major products
Arginine	Glutamic semialdehyde
Cysteine	-S-S- disulfide cross-links
Histidine	2-Oxohistidine, aspartate, asparagine
Leucine	Hydroxyleucine
Lysine	2-Aminoadipic semialdehyde
Methionine	Methionine sulfoxide
Phenylalanine	o-Tyrosine
Proline	Glutamate, pyroglutamate, cis/trans-4-hydroxyproline,
	2-pyrrolidone, glutamic semialdehyde, γ-aminobutyric acid
Threonine	2-Amino-3-ketobutyric acid
Tryptophane	N-Formylkynurenine
Tyrosine	Tyr-Tyr cross-links

FIG. 2. Structure of methionine and its oxidation products.

sulfhydryl (cysteine) or basic amino acids (histidine, lysine) (42, 116). Increased exposure of proteins and lipoproteins to lipid-derived aldehydes may be representative of oxidant stress in various pathologies. Only a few studies have directly addressed the quantitative damage in complex systems. However, oxidized amino acids in proteins are emerging as attractive candidate markers of oxidant stress (63).

ROS interaction with nitric oxide (NO) results in the generation of a number of radical and nonradical reactive nitrogen species (RNS).

These aspects are the subject of important developments in cell signaling and protein modifications (6, 25, 30). NO itself and peroxynitrite (ONOO⁻) are also able to modify proteins and amino acid residues oxidatively. The interaction of NO with O_2^{-} seems to attenuate the beneficial actions of NO, such as stimulation of soluble guanylate cyclase and effects on mitochondrial respiration. The resulting oxidation products, such as 3-nitrotyrosine, may have important biological functions (39). The most potent actions of ONOO⁻ and RNS appear to be

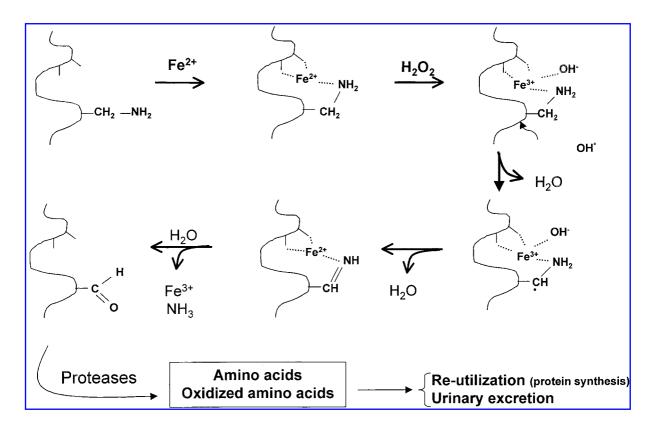


FIG. 3. Proposed mechanism for the metal-catalyzed oxidation of an ϵ -amino group of a lysine residue in a protein and its degradation (redrawn from 109).

thiol modifications. RNS may also participate in cellular signaling processes through interactions with prostaglandins. An example can be illustrated by the inactivation of prostaglandin GI₂ synthase as a result of nitration by ONOO of a tyrosine residue (128). HOCl produced by neutrophils to kill foreign organisms can also be regarded as a reactive species. Hypochlorite-oxidized proteins that contain 3-chlorotyrosine probably originating from oxidized apolipoprotein B after interaction with myeloperoxidase have been detected in atherosclerotic plaques (62).

BIOLOGICAL SIGNIFICANCE OF PROTEIN OXIDATION

The biological significance of ROS-induced damage to proteins has important physiological implications. Numerous studies have shown that such modifications are relevant for protein activity, cell functioning, receptor-mediated processes, and degradation (30). These modifications can be related to induced changes in electronegativity, loss of tryptophan, bityrosine formation (Fig. 4), or the appearance of disulfide cross-links. These modifications may occur at the intra- or intermolecular level and may favor protein aggregation. Other modifications may also result from indirect alterations by lipid oxidation products.

Various biological roles have been proposed for ROS-mediated damage to proteins. Some are key elements of physiological processes like protein turnover and regulation, whereas others are implicated in pathological disorders. The precise metabolic fate of oxidized proteins requires specific studies. However, these types of studies are difficult to conduct because oxidative modifications of proteins might be complex. Indeed, experiments carried out in simple oxidative systems, in which only the amino acid residues are oxidized, indicated that oxi-

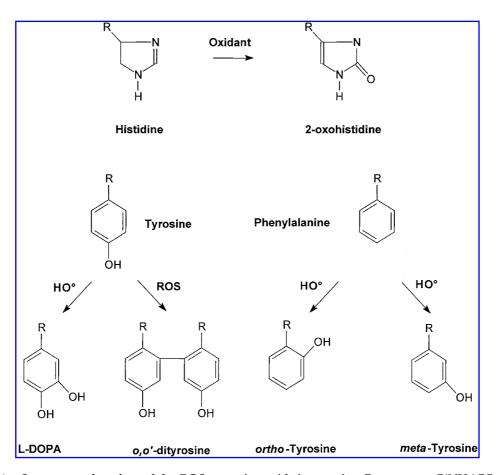


FIG. 4. Some examples of attack by ROS on amino acids in proteins. R represents C(NH₂)COOH.

dized proteins are usually sensitive to proteolytic attack by most proteases (27, 30, 83). However, in more complex systems, in which proteins might be modified by products of lipid and carbohydrate oxidation, the range of modifications is wider (42, 116, 120). In these latter situations, which are more analogous to what is encountered in biology, proteins are more heavily modified and show decreased susceptibility to proteases (27). This has been described for aldehyde-modified apolipoprotein B found in oxidized low-density lipoproteins (LDL), which was poorly degraded compared with native LDL. Various aldehydes originate from lipid oxidation products, including reactive aldehydes (malondialdehyde, 4-hydroxynonenal) (42). Other protein-modifying agents are produced by nonenzymatic glycation, defined as the chemical union of glucose to protein. Several not entirely defined products are formed beginning with a Schiff base and undergoing a series of subsequent reactions and/or rearrangements until formation of advanced species corresponding to brown or fluorescent pigments called advanced glycation end products (AGE) (15). These products accumulate in long-life proteins, such as basal membrane proteins (collagen or lens proteins). Some examples include alkyl-formyl-diglycoxylpyrrole (a pentose-derived cross-link between arginine and lysine), pyrroline, carboxymethyllysine, and pentosidine. AGE elicit a wide range of cell-mediated responses that are thought to be largely induced through an AGE-specific cell-surface receptor (121). Although precise information about the rates of catabolism, pool sizes, and half-lives of native and oxidized/glucoxidized proteins is still needed, accumulation might be the fate of highly oxidized proteins (54, 73). It has been suggested that macroxyproteinases or proteasomes are part of the selective degradation of oxidatively modified proteins (83, 95). Actually, the various data available show that mild oxidation of proteins leads to an increased sensitivity to proteases, whereas extensive oxidation is associated with resistance to proteolysis. These differences could be explained by the fact that proteases might have a limited access to strongly oxidized proteins because of marked conformational changes due to denaturation

and/or aggregation (30). Elevated levels of protein carbonyls and various compounds resulting from oxidative modification of amino acids have been reported in aging, neurodegenerative diseases, atherosclerosis, and diabetes (7, 83). However, the increased susceptibility of mildly oxidized proteins to proteolytic degradation may explain an increased urinary excretion of oxidized amino acids (69). As a matter of fact, normal amino acids are known to be reutilized but it is still not known whether oxidized amino acids are processed for protein biosynthesis at a normal rate. Alternatively, when oxidatively induced changes are still reversible, damaged proteins may undergo direct repair and thus be spared from proteolytic degradation.

SPECIFIC ROLE OF SULFUR CONTAINING AMINO ACID RESIDUES IN PROTEINS

Thiols

As discussed above, cysteine and methionine belong to the most easily oxidized amino acids. Conversion of SH groups into disulfides is one of the early steps of protein oxidation (81, 109, 115). Within a protein, oxidation of two nearby cysteine residues results in the formation of a disulfide bridge producing a more rigid protein. Disulfide bonds can also form between two proteins, and it can result in a very complex structure at the supramolecular level with several proteins cross-linked together. It is easily understandable that the dynamic properties of cell membranes may strongly be altered when such processes occur. The changes can be monitored using the sulfhydrylselective spin label MAL-6 (2,2,6,6-tetramethyl-4-maleimidopiperidine-1-oxyl), which covalently binds only to SH groups on the proteins (108). The difference in spin-label motion indicates modifications of protein interactions. Other modifications may also exist when lowmolecular-weight thiols, such as reduced free SH-containing amino acids or glutathione, are linked to proteins. This process is called protein thiolation (115) and appears to be favored by oxidant stress (Fig. 5). Not every SH group

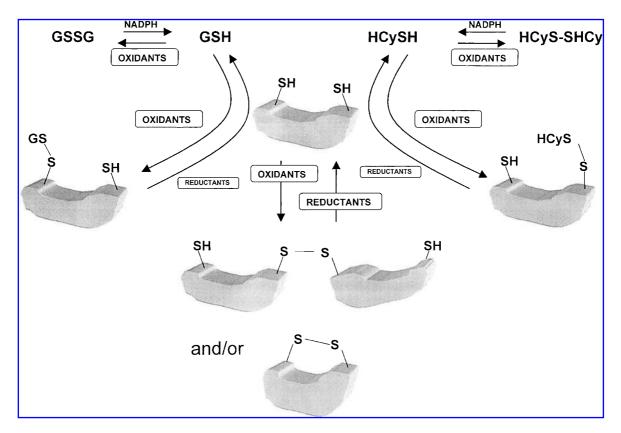


FIG. 5. Various proposed oxidation and reduction processes resulting in protein thiolation/dethiolation. GSH and GSSG: reduced and oxidized glutathione, respectively; HCySH and HCyS-SHCy: homocysteine and homocystine, respectively; Reductants: GSH, glutaredoxin, thioredoxin. Oxidants: O₂-', H₂O₂, HO', HOCl, RO', ROO', NO', ONOO-.

of a protein may react with the same efficacy and, obviously, the reactivity depends on the local environment and/or whether the cysteine residue is buried or exposed. An important feature of the oxidation of thiol groups is its reversibility in the presence of appropriate reductants. Reduced glutathione, exogenous dithiothreitol (in vitro), or enzymes are involved in this dethiolation process. Much of our current understanding of the thiolation/ dethiolation process as an early response to oxidative stress arose from experiments carried out with reactive thiol groups (96). Such a limited oxidation could be transient, and it could be involved in the regulation of enzyme activity and in the maintenance of protein folding/conformation. One example can be illustrated by the complete inactivation of creatine kinase by a single S-thiolation of each subunit (115). However, there is no evidence that this process could be an enzyme-driven process, suggesting that direct reaction with oxidizing materials prevails. Moreover, oxidation-induced changes of protein conformation may bring a formerly distant thiol group in the proper vicinity to react quickly. For the dethiolation reaction, available data suggest that dithiol redox proteins, such as thioredoxin or glutaredoxin, are involved (55, 93, 115).

Homocysteine (Hcy)

Among the sulfhydryl amino acids, it is of importance to consider specifically the case of Hcy. Hyperhomocysteinemia, caused by the accumulation of this amino acid in plasma, is recognized as a strong and independent risk factor for cardiovascular disease, as well as venous and arterial thrombosis (11, 20, 38, 53, 97). Experimental evidence has established that iatrogenic or nutritional folate deficiency is also related, through hyperhomocysteinemia, to the thrombotic side of vascular disease (35–37). In rats, it has been recently demonstrated that long-term folate deficiency favors the occurrence of a prethrombotic state consequent to an

oxidative stress (35). The imbalance of the redox status mediated by folate deficiency has been characterized by various indices of oxidation, such as an increase in end products of lipid oxidation (thiobarbituric acid reactive substances, conjugated dienes, hydroperoxides), changes in fatty acid composition with a decrease in polyunsaturated fatty acids (mostly n-3 fatty acids), and a lower antioxidant defense as shown using our free radical-mediated erythrocyte hemolysis test (8, 34, 87). The prothrombotic and prooxidant effects of folate deficiency have been linked to a moderate increase in circulating Hcy levels. When methionine loading is used as the sole source of Hcy, plasma Hcy was demonstrated to be correlated both to lipoperoxidation markers and to platelet aggregation, as well as to the biosynthesis of the potent proaggregant thromboxane (36).

A high proportion of HcySH (>50%) is found bound to plasma proteins either by disulfide bonds to cysteine residues or by peptide bonds to lysine residues. Experimental evidence is still needed to increase our knowledge of the consequence of thiolation of proteins by Hcy. However, it has been proposed that in aging tissues the oxidative condition of proteins and the loss of catalytic activity of enzymes may be related to changes in Hcy metabolism (74, 82). Hcy, or its derivatives, may combine with lysine groups of elastin, preventing desmosine formation. The thiolation of the apolipoprotein B of LDL by Hcy can promote oxidation of the lipoprotein. It is now well documented that oxidized LDL strongly enhances cholesterol accumulation in macrophages and that this results in atherogenesis (43, 111). These effects may depend on the type of adduct formed. When the reaction involves the thiol group of Hcy and protein, the result is a decrease in the antioxidant properties of the thiolated protein. Conversely, it has been also demonstrated that when Hcy reacted in its thiolactone form with LDL, the resulting homocystamide–LDL adduct was more resistant to oxidation, possibly due to an increase in thiols (47) according to Scheme 1. However, the precise conditions leading to thiolation or homocystamide adduct are unknown.

Methionine oxidation

As mentioned above, methionine can be oxidized by a great variety of oxidants such as H_2O_2 , hydroxyl radicals, hypochlorite, chloramine, and ONOO $^-$. The resulting oxidation product obtained in the first step is methionine sulfoxide (12). Oxidation to sulfone, which is the next step, is only obtained under drastic conditions not usually occurring in biological systems. It is noteworthy that methionine sulfoxide is quickly reversed back to methionine with mild reductants, whereas sulfone formation is biologically irreversible.

There are very few studies reporting on the occurrence of methionine sulfoxide in proteins during oxidation-related diseases. This could be due in part to three main factors all related to the difficulty of analyzing this modified amino acid. First, routine procedures, such as amino acid analysis and automated Edman se-

quencing, usually do not identify oxidized methionine because of its reduction back to methionine. Second, oxidation of methionine residues does not lead to formation of a carbonyl group that could react with the widely utilized carbonyl assay for oxidatively modified proteins. Finally, the only technique available at present is rather cumbersome. As CNBr cleaves peptide bonds on the carboxyl side of methionine only, yielding homoserine, and does not cleave at methionine sulfoxide, the technique of simultaneous sequencing of the CNBr peptides allows determination of the status of most of the methionine residues (48).

Several examples of oxidation of methionine residues in proteins have been reported. These data confirm the importance of this process in protein function (12, 122). Some studies concern the H₂O₂-mediated oxidation of methionine residues in L12 ribosomal protein resulting in impaired protein synthesis (12). This activity can be reactivated by reduction with mercaptoethanol or by methionine sulfoxide reductase (12, 13, 16). Other examples come from studies on activity of antiproteases. It has been reported that the antiproteolytic activity of human α_2 -macroglobulin was inhibited by oxidation and that methionine and tryptophan play critical roles in controlling its structure and function (94). Similar observations were reported for α_1 -proteinase inhibitor present in blood plasma and other body fluids, which inhibits a variety of serine proteases. This protein contains eight methionyl residues. Treatment with oxidants leads to oxidation of two surfaceexposed methionine to methionine sulfoxide, resulting in a drastic loss of the inhibition of elastase activity (for review, see 44). No cleavage occurs, and no other amino acid residue is affected. The biological significance of the impairment of the activity of α_1 -proteinase inhibitor has been related to the frequent occurrence of emphysema in cigarette smokers due to the oxidative stress induced by cigarette smoke and activated leukocytes in the lungs (9). Another example comes from data dealing with activation of the fifth component of human complement (C5), which normally occurs through the action of convertases formed from other complement components (122). It was found that oxidants lead to oxidation of methionine without peptide cleavage. Whereas the physiological activation of C5 is restricted to targets, oxidant-activated C5 bearing oxidized methionine is able to diffuse and might attack cells far from its origin. This results in subsequent release of anaphylatoxin activity.

Others examples of the importance of methionine oxidation may also be found in proteins from various origins. It has been shown that the content of the oxidized residue measured in lens proteins increases with age (64, 122). Apolipoprotein AI, which is a main protein component of high-density lipoproteins, is involved in the clearance of cholesterol back to the liver (reverse cholesterol transport; see 98 for review). Although other components, such as phospholipids and oxysterols, are known to be of importance (50), it has been shown that oxidation of apolipoprotein AI by H₂O₂ leads to reduced binding capacity for lipids. It has been proposed that the introduction of oxidized methionine residues in apolipoprotein AI resulted in change in secondary structure due to modification of its hydrophobicity (1). All these selected examples clearly show that the integrity of methionine has to be maintained for an optimal cell functioning and that oxidation of methionine may be related to the onset of pathological diseases.

Methionine sulfoxide reductase

Until recently, the occurrence of oxidized methionine residues in proteins was merely considered as part of their denaturation process. Oxidation of methionine residues in proteins, which can be reversed enzymatically, may suggest that oxidation of methionine and its reversal may serve to regulate the various functions of proteins (12, 122). Although only found so far at the intracellular level, this widespread enzyme appears capable of reducing either the free or the protein-bound form of methionine sulfoxide. This enzyme has been cloned from bacteria (14, 91), plants (99), and bovine adrenal gland (79). Thus, the hypothesis concerning the role of methionine sulfoxide reductase as part of a repair system resulting in free radical detoxification was recently put forward in several articles from the groups of Brot and Stadtman (see Scheme 2) (70, 78, 79). This idea has recently been strengthened by the finding that overexpression of methionine sulfoxide reductase in yeast and human T cells provided them with high resistance to H₂O₂mediated oxidative stress (78). Recent findings also suggest that methionine oxidation might be a regulator of cellular excitability. The functional properties of a transient A-type potassium channel expressed in Xenopus oocytes were impaired by oxidizing a critical methionine residue (22). The channel protein was protected from oxidation by antioxidants, but most importantly by coexpression of methionine sulfoxide reductase (23).

MT AS AN INTRACELLULAR ANTIOXIDANT

MTs are ubiquitous, low-molecular weight proteins (6000–7000 Da) with still unclear functions (66, 113, 119). They were characterized by their unusually high content in thiols. All vertebrates contain two or more distinct MT forms designed MT-1 to MT-4. In mammals, MT-1 and MT-2 are found in all organs, whereas MT-3 is most abundant in brain. First identified as cadmium-binding proteins, MTs are known to form high-affinity complexes with other trace metals, including mercury, silver, zinc and copper. As these metals are also able to induce MT

expression (mostly MT-1 and MT-2), a critical role has been proposed for MTs in intracellular metal ion homeostasis. The 20 cysteines of a total of 61 amino acids of the MT molecules form two thiolate clusters involved in metal binding. Most of the past studies indicated that extracellular Cd-MT complexes are toxic, especially for the kidney. These effects are in contrast with the induction of MT, which allows cells to sequester intracellular Cd in a nontoxic form (113). Thus intracellular MT appears to be protective, whereas extracellular MT is harmful especially during metal exposure. The large number of free thiols in MT suggests that these proteins might be subjected to sulfhydryl oxidation cycles. It has been demonstrated that oxidation with H₂O₂ and various free radicals induces metal release that might become available for Fenton reactions (45, 89). These findings allowed a role for MT in antioxidant defense to be proposed (102). This idea is in full agreement with previous findings that MT is a redox-sensitive gene whose expression is elevated by oxidative stress. Furthermore, an enhanced sensitivity to oxidant injury has been shown in MT-null cells and mice sensitive to Cd and oxidant exposure (67). Conversely, overexpression of MT decreases sensitivity to oxidant injury (86, 104).

The precise mechanism of the antioxidant activity of MT is still unclear. Data resulting from *in vitro* experiments clearly indicated that MT scavenges $O_2^{-\bullet}$ and various types of radicals

such as hydroxyl, phenoxyl, and NO radicals (102–104). It is likely that this activity could be due to the high content of cysteine residues involved in thiolate clusters. It is considered that MT is indeed an expandable target for free radicals and that it may work in conjunction with an unknown regenerating system. However, the capability of MT to inhibit both metal-dependent and -independent lipid peroxidation would suggest that the antioxidant role of MT does not rely only on its interaction with metals.

ALBUMIN AS A CIRCULATING ANTIOXIDANT

Albumin

With 585 amino acids and a molecular weight of 69 kDa, this highly soluble protein that does not contain carbohydrate is present in human plasma at normal concentrations between 35 and 50 g/L. Albumin has a number of important physiological and pharmacological functions, such as a transporter for metals, fatty acids, cholesterol, NO, bile pigments, and drugs. It is a key element of the regulation of osmotic pressure and fluid distribution between different compartments. In normal conditions, its half-life is \sim 20 days and its plasma concentration represents an equilibrium not only between its synthesis by the liver and its catabolism, but also its transcapillary escape. As indicated above, albumin may actually represent the major and predominant antioxidant in plasma, a body compartment known to be exposed to continuous oxidative stress.

Epidemiological data

Most of the epidemiological studies have established an inverse relationship between serum albumin levels and mortality risk. In diseased populations, as well as in the general population, it has been estimated that the odds of death increase by $\sim 50\%$ for each 2.5 g/L decrement in the initial albumin level (for review, see 52). This association holds also for cardiovascular disease after adjustment for the usual risk factors (72, 85). Variation in albumin concentration may reflect variation in nutritional state. In fact, only a small number of fac-

tors are known to result in variation in serum albumin concentration. Beside analbuminemia, a rare congenital disease (24, 77), the main pathological situation known to lower albumin concentration is the nephrotic syndrome, which is the subject of many studies (61, 125). In addition, it has been reported that serum albumin decreases with age and cigarette smoking (85).

Modifications of albumin structure

Alteration of the structure of albumin may result in modifications of its biological properties. As albumin is capable of reacting with most oxygenated species, this may lead to some oxidative-induced changes, which can explain why albumin exists in an equilibrium between its reduced and oxidized form. Although in normal conditions the reduced form predominates, this equilibrium could shift depending on pathological states and during aging (41). Products of lipid peroxidation, such as reactive aldehydes, may also react with lysine or arginine residues, resulting in albumin alteration. These modifications, together with structural and functional alterations due to increased glycation, could occur in insulin-dependent diabetes mellitus, which is one of the pathological conditions associated with early occurrence of vascular complications (126). The binding of glucose to albumin typically occurs in vivo and is known to involve the nonenzymatic covalent attachment of glucose to a lysine side chain. Approximately 6-10% of the albumin in normal human serum is modified by nonenzymatic glycosylation (106). This proportion typically increases between two and three times in hyperglycemia (127). Moreover, diabetic patients exhibit elevated levels of iron and copper ions, which, in the presence of glycated proteins, have been shown in vitro to generate free radicals (29). These highly reactive species are able to induce oxidative degradation of protein in vitro (83). If the biosynthesis process is fully functional, damaged albumin may be degraded by proteolysis and quickly replaced. It has been calculated that in normal conditions, \sim 3 g of albumin must be synthesized daily just to keep the basal concentration (56).

In a recent work, the antioxidant activity of

bovine serum albumin (BSA) has been addressed (10). By using different approaches, it has been found that first, albumin markedly delayed the copper-initiated oxidation of LDL as monitored by the appearance of conjugated dienes (Fig. 6), by changes in electrophoretic mobility and by formation of end products of fatty acid oxidation, such as thiobarbituric acid reactive substances. Second, albumin offered a significant protection against free radical-mediated hemolysis. These antioxidant effects were found to be concentration-dependent, which is consistent with the idea of a beneficial effect of high albumin levels in humans. However, besides its concentration, the integrity of the albumin molecule might be a key determinant of its activity. When BSA was preincubated in the presence of various glucose concentrations, it progressively lost its antioxidant properties as regards measured indices of the propensity of LDL to oxidize. Under our conditions, data suggest that only minor modifications of the BSA molecule and no large damage occurred following incubation with glucose and with free radicals. Oxidation of BSA by free radicals decreased its thiol content. Of the 35 cysteines of the BSA molecule, 34 are engaged in 17 S-S-bonded cystines, leaving only Cys³⁴ available for reactions (84). As isolated from plasma, about one-third of the albumin molecules carry a mixed disulfide picked up in plasma (cysteine, reduced glutathione, Hcy) on this cysteine. When this cysteine residue has been selectively blocked by in vitro treatment with N-ethymaleimide, albumin lost most of its thiol groups, and this resulted in a significant reduction in its antioxidant properties as assessed by free radicalmediated hemolysis (Fig. 7). However, the significant antioxidant activity remaining in the thiol-blocked BSA indicated that other amino acid residues (Tyr, Trp, Met) participated in the total activity. As suggested by the group of Davies (28, 65) using electron paramagnetic resonance spin trapping with BSA (and other proteins), thiols may work either as a sink for radicals, thus protecting the protein from entire denaturation, or as agents transferring the damage to other sites, such as the peptide backbone. These data obtained with albumin as an experimental model confirm that protein-toprotein damage transfer and protein-oxidation reactions may readily occur. This may explain why it was found that, in some instances, BSA not only lost its antioxidant effect, but became prooxidant (Fig. 6).

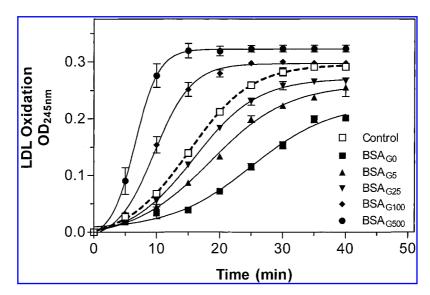


FIG. 6. Effects of glucose- and oxidation-modified BSA on the Cu^{2+} -induced LDL oxidation. Oxidation of LDL was started by adding $40~\mu M$ CuSO₄ and monitored by the absorbance of the conjugated dienes in the absence (control, dashed line) or presence of various BSA preparations (2 mg/ml). BSAG_x refers to BSA incubated for 60 days with x mM glucose as described (10). This figure is reproduced with the permission of FASEB J.

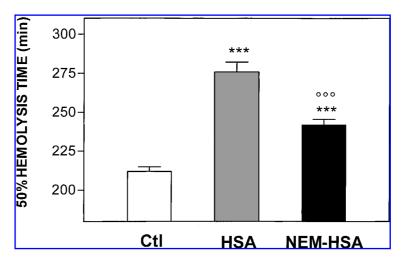


FIG. 7. Effects of modified albumin on free radical-induced hemolysis. Experiments, performed as in Fig. 1, were carried out with native human serum albumin (HSA) and with N-ethylmaleimide (NEM)-modified HSA (2 mg/ml), which blocked accessible thiols measured as described (10) using Ellman's assay (40). Corresponding thiol values (n = 3) were 4.3 ± 1.0 and 1.3 ± 0.4 pmol/mg of protein for native HSA and NEM-HSA, respectively. Results are expressed as 50% hemolysis time in minutes.

CONCLUDING REMARKS

Oxidative damage to proteins may be of particular importance for severeal reasons. First, it affects the normal function of proteins, e.g., receptor and enzyme activities, transport proteins, coagulation process, peptide hormones (4, 46, 92, 101, 105, 107, 112, 114, 117). Second, the appearance of ROS-induced epitopes on proteins may contribute to the generation of new antigens that can provoke cell-mediated and immune responses. Finally, the loss of the normal function of proteins may result in secondary damage to other biomolecules, e.g., inactivation of DNA repair enzymes, changes in the rates of biosynthesis of mediators (cyclic nucleotides, prostaglandins), and receptor recycling. The important role of oxidized proteins is supported by the growing number of reports indicating that tissue concentrations of oxidized amino acids are increased in pathological situations and in experimental models (30, 68, 69). The important role of oxidized proteins is further reinforced by the fact that thiol repletion achieved with albumin infusion to patients has been suggested to restore the antioxidant balance (90). However, further work is needed to precisely document ex vivo the occurrence of oxidized proteins using standardized techniques and to confirm the validity of specific markers, in particular after action of antioxidants. The specific role of methionine, which could act not only as an endogenous antioxidant, but also as a regulatory molecule, has to be more thoroughly considered due to the reversibility between its reduced and oxidized forms. The functionality of methionine sulfoxide reductase needs to be clearly established in future research. Mild oxidant stress can result in up-regulation of certain antioxidants to restore the balance between free radical attack and defense; however, repeated severe oxidant stress produces major interdependent alterations that can lead to chronic disease.

ACKNOWLEDGMENTS

Work performed by the authors was supported by the Institut National de la Santé et de la Recherche Médicale (INSERM), the Conseil Régional de Bourgogne, the Université de Bourgogne, and a grant from ARCOL. The helpful comments of Dr. Lise Bernier (IRCM) were kindly appreciated. E.B. is supported by a fellowship from the Ministère de l'Education Nationale, de l'Enseignement Supérieure et de la Recherche.

ABBREVIATIONS

AGE, advanced glycation end products; BSA, bovine serum albumin; Hcy, homocysteine; H₂O₂, hydrogen peroxide; LDL, low-density lipoproteins; MT, metallothionein; NO, nitric oxide; O₂^{-•}, superoxide anion; ONOO⁻, peroxynitrite; RNS, reactive nitrogen species; ROS, reactive oxygen species; SOD, superoxide dismutase.

REFERENCES

- Anantharamaiah GM, Hughes TA, Iqbal MP, Gawish A, Neame PJ, Medley MF, and Segrest JP. Effect of oxidation on the properties of apolipoproteins A-I and A-II. Adv Exp Med Biol 281: 401–408, 1990.
- Aruoma OI. Nutrition and health aspects of free radicals and antioxidants. Food Chem Toxicol 32: 671–683, 1994.
- 3. Aruoma OI. Free radicals, oxidative stress, and antioxidants in human health and disease. *J Am Oil Chem Soc* 75: 199–212, 1998.
- 4. Asahi M, Fujii J, Suzuki K, Seo HG, Kuzuya T, Hori M, Tada M, Fujii S, and Taniguchi N. Inactivation of glutathione peroxidase by nitric oxide—implication for cytotoxicity. *J Biol Chem* 270: 21035–21039, 1995.
- Aslan R, Sekeroglu MR, Gültekin F, and Bayiroglu F. Blood lipoperoxidation and antioxidant enzymes in healthy individuals: relation to age, sex, habits, life style and environment. *J Environ Sci Health [A]* 32A: 2101–2109, 1997.
- Beckman JS, Chen J, Ischiropoulos H, and Crow JP. Oxidative chemistry of peroxynitrite. *Methods Enzymol* 233: 229–240, 1994.
- 7. Berlett BS, and Stadtman ER. Protein oxidation in aging, disease, and oxidative stress. *J Biol Chem* 272: 20313–20316, 1997.
- 8. Blache D, and Prost M. Free radical attack: biological test for human resistance capability. In: *Proceedings of the IX College Park Colloquium on Chemical Evolution: A Lunar-Based Chemical Analysis Laboratory (LBCAL)*, edited by Ponnamperuma C, and Gehrke CW. Washington, DC, 1992, pp. 82–98.
- 9. Blache D, Bouthillier D, and Davignon J. Acute influence of smoking on platelet behaviour, endothelium and plasma lipids and normalization by aspirin. *Atherosclerosis* 93: 179–188, 1992.
- 10. Bourdon E, Loreau N, and Blache D. Glucose and free radicals impair the antioxidant properties of serum albumin. *FASEB J* 13: 233–244, 1999.
- Boushey CJ, Beresford SAA, Omenn GS, and Motulsky AG. A quantitative assessment of plasma homocysteine as a risk factor for vascular disease—probable benefits of increasing folic acid intakes. *JAMA* 274: 1049–1057, 1995.

- 12. Brot N, and Weissbach H. Biochemistry and physiological role of methionine sulfoxide residues in proteins. *Arch Biochem Biophys* 223: 271–281, 1983.
- Brot N, Weissbach L, Werth J, and Weissbach H. Enzymatic reduction of protein-bound methionine sulfoxide. *Proc Natl Acad Sci U S A* 78: 2155–2158, 1981.
- 14. Brot N, Rahman MA, Moskovitz J, and Weissbach H. *Escherichia coli* peptide methionine sulfoxide reductase: cloning, high expression, and purification. *Methods Enzymol* 251: 462–470, 1995.
- Brownlee M, Cerami A, and Vlassara H. Advanced glycosylation end products in tissue and the biochemical basis of diabetic complications. N Engl J Med 318: 1315–1321, 1988.
- Caldwell P, Luk DC, Weissbach H, and Brot N. Oxidation of methionine residues of *Escherichia coli* ribosomal protein L12 decreases the protein's biological activity. *Proc Natl Acad Sci U S A* 75: 5349–5352, 1978.
- 17. Cao G, Verdon CP, Wu AHB, Wang H, and Prior RL. Automated assay of oxygen radical absorbance capacity with the COBAS FARA II. *Clin Chem* 41: 1738–1744, 1995.
- Cao GH, and Prior RL. Comparison of different analytical methods for assessing total antioxidant capacity of human serum. Clin Chem 44: 1309–1315, 1998.
- 19. Cao GH, Giovanoni M, and Prior RL. Antioxidant capacity in different tissues of young and old rats. *Proc Soc Exp Biol Med* 211: 359–365, 1996.
- Cattaneo M. Hyperhomocysteinemia, atherosclerosis and thrombosis. *Thromb Haemost* 81: 165–176, 1999.
- 21. Church OT, and Pryor WA. Free radical chemistry of cigarette smoke and its toxicological implications. *Environ Health Perspect* 64: 111–126, 1985.
- 22. Ciorba MA, Heinemann SH, Brot N, Weissbach H, and Hoshi T. Modulation of potassium channel function by methionine oxidation and reduction. *Proc Natl Acad Sci U S A* 93: 15306–15340, 1999.
- 23. Ciorba MA, Heinemann SH, Weissbach H, Brot N, and Hoshi T. Regulation of voltage-dependent K⁺ channels by methionine oxidation: effect of nitric oxide and vitamin C. FEBS Lett 442: 48–52, 1999.
- 24. Dammaco F, Miglietta A, D'Addabbo A, Fratello A, Moschetta R, and Bonomo L. Analbuminemia: report of a case and review of the literature. *Vox Sang* 39: 153–161, 1980.
- 25. Darley-Usmar V, and Halliwell B. Blood radicals—reactive nitrogen species, reactive oxygen species, transition metal ions, and the vascular system. *Pharm Res* 13: 649–662, 1996.
- 26. Davies KJA. Oxidative stress: the paradox of aerobic life. *Biochem Soc Symp* 61: 1–31, 1995.
- 27. Davies KJA, Lin SW, and Pacifici RE. Protein damage and degradation by oxygen radicals. IV. Degradation of denatured protein. *J Biol Chem* 262: 9914–9920, 1987.
- 28. Davies MJ, Gilbert BC, and Haywood RM. Radical-

- induced damage to bovine serum albumin: role of the cysteine residue. *Free Radic Res Commun* 18: 353–367, 1993.
- 29. Dean RT, Hunt JV, Grant AJ, Yamamoto Y, and Niki E. Free radical damage to proteins: the influence of the relative localization of radical generation, antioxidants, and target proteins. *Free Radic Biol Med* 11: 161–168, 1991.
- 30. Dean RT, Fu SL, Stocker R, and Davies MJ. Biochemistry and pathology of radical-mediated protein oxidation. *Biochem J* 324: 1–18, 1997.
- 31. De Zwart LL, Meerman JHN, Commandeur JNM, and Vermeulen NPE. Biomarkers of free radical damage applications in experimental animals and in humans. *Free Radic Biol Med* 26: 202–226, 1999.
- 32. Diplock AT, Charleux JL, Crozier-Willi G, Kok FJ, Rice-Evans C, Roberfroid M, Stahl W, and Viña-Ribes J. Functional food science and defence against reactive oxidative species. *Br J Nutr* 80: S77–S112, 1998.
- 33. Doetsch PW, Helland DE, and Haseltine WA. Mechanism of action of a mammalian DNA repair endonuclease. *Biochemistry* 25: 2212–2220, 1986.
- Durand P, and Blache D. Enhanced platelet thromboxane synthesis and reduced macrophage-dependent fibrinolytic activity related to oxidative stress in oral contraceptive-treated female rats. *Atherosclerosis* 121: 205–216, 1996.
- 35. Durand P, Prost M, and Blache D. Pro-thrombotic effects of a folic acid deficient diet in rat platelets and macrophages related to elevated homocysteine and decreased n-3 polyunsaturated fatty acids. *Atherosclerosis* 121: 231–243, 1996.
- 36. Durand P, Lussier-Cacan S, and Blache D. Acute methionine load-induced hyperhomocysteinemia enhances platelet aggregation, thromboxane biosynthesis and macrophage-derived tissue factor activity in rats. *FASEB J* 11: 1157–1168, 1997.
- Durand P, Prost M, and Blache D. Folic acid deficiency enhances oral contraceptive-induced platelet hyperactivity. *Arterioscler Thromb Vasc Biol* 17: 1939–1946, 1997.
- 38. Durand P, Prost M, and Blache D. Folate deficiency and cardiovascular pathologies. *Clin Chem Lab Med* 36: 419–429, 1998.
- 39. Eiserich JP, Vossen V, O'Neill CA, Halliwell B, Cross CE, and Van der Vliet A. Molecular mechanisms of damage by excess nitrogen oxides: nitration of tyrosine by gas-phase cigarette smoke. *FEBS Lett* 353: 53–56, 1994.
- 40. Ellman GL. Tissue sulfhydryl groups. *Arch Biochem Biophys* 82: 70–77, 1959.
- 41. Era S, Kuwata K, Imai H, Nakamura K, Hatashi T, and Sogami M. Age-related change in the redox state of human serum albumin. *Biochim Biophys Acta* 1247: 12–16, 1998.
- 42. Esterbauer H, Schaur RJ, and Zollner H. Chemistry and biochemistry of 4–hydroxynonenal, malonaldehyde and related aldehydes. *Free Radic Biol Med* 11: 81–128, 1991.

- Esterbauer H, Gebicki J, Puhl H, and Jürgens G. The role of lipid peroxidation and antioxidants in oxidative modification of LDL. *Free Radic Biol Med* 13: 341–390, 1992.
- 44. Evans MD, and Pryor WA. Cigarette smoking, emphysema, and damage to α₁-proteinase inhibitor. *Am J Physiol* 266: L593–L611, 1994.
- Fabisiak JP, Tyurin VA, Tyurina YY, Borisenko GG, Korotaeva A, Pitt BR, Lazo JS, and Kagan VE. Redox regulation of copper-metallothionein. *Arch Biochem Biophys* 363: 171–181, 1999.
- Faure P, Lafond J-L, Coudray C, Rossini E, Halimi S, Favier A, and Blache D. Zinc prevents the structural and functional properties of free radical treated-insulin. *Biochim Biophys Acta* 1209: 260–264, 1994.
- 47. Ferguson E, Hogg N, Antholine WE, Joseph J, Singh RJ, Parthasarathy S, and Kalyanaraman B. Characterization of the adduct formed from the reaction between homocysteine thiolactone and low-density lipoprotein: antioxidant implications. *Free Radic Biol Med* 26: 968–977, 1999.
- 48. Fliss H, Weissbach H, and Brot N. Oxidation of methionine residues in proteins of activated human neutrophils. *Proc Natl Acad Sci U S A* 80: 7160–7164, 1983.
- Fridovich I. Superoxide radical and superoxide dismutases. *Annu Rev Biochem* 64: 97–112, 1995.
- Gesquière L, Loreau N, and Blache D. Impaired cellular cholesterol efflux by oxysterol-enriched high density lipoproteins. *Free Radic Biol Med* 23: 541–547, 1997.
- 51. Girodon F, Blache D, Monget AL, Lombart M, Brunet-Lecompte P, Arnaud J, Richard MJ, and Galan P. Effect of two year supplementation with low dose antioxidant vitamins and/or minerals in elderly subjects on levels of nutrients and on antioxidant defense parameters. *J Am Coll Nutr* 16: 357–365, 1997.
- 52. Goldwasser P, and Feldman J. Association of serum albumin and mortality risk. *J Clin Epidemiol* 50: 693–703, 1997.
- 53. Graham IM, Daly LE, Refsum HM, Robinson K, Brattström LE, Ueland PM, Palma-Reis RJ, Boers GHJ, Sheahan RG, Israelsson B, Uiterwaal CS, Meleady R, McMaster D, Verhoef P, Witteman J, Rubba P, Bellet H, Wautrecht JC, De Valk HW, Lúis ACS, Parrot-Roulaud FM, Tan KS, Higgins I, and Garcon D. Plasma homocysteine as a risk factor for vascular disease—the European concerted action project. *JAMA* 277: 1775–1781, 1997.
- Grant AJ, Jessup W, and Dean RT. Accelerated endocytosis and incomplete catabolism of radical-damaged protein. *Biochim Biophys Acta* 1134: 203–209, 1992.
- 55. Gravina SA, and Mieyal JJ. Thioltransferase is a specific glutathionyl mixed disulfide oxidoreductase. *Biochemistry* 32: 3368–3376, 1993.
- 56. Halliwell B. Albumin—an important extracellular antioxidant? *Biochem Pharmacol* 37: 569–571, 1988.

- 57. Halliwell B. Antioxidants in human health and disease. *Annu Rev Nutr* 16: 33–50, 1996.
- 58. Halliwell B. Oxidative stress, nutrition and health. Experimental strategies for optimization of nutritional antioxidant intake in humans. *Free Radic Res* 25: 57–74, 1996.
- 59. Halliwell B, and Cross CE. Oxygen-derived species: their relation to human disease and environmental stress. *Environ Health Perspect* 102 (Suppl 10): 5–12, 1994.
- 60. Harris ED. Regulation of antioxidant enzymes. *FASEB J* 6: 2675–2683, 1992.
- 61. Harris RC, and Ismail N. Extrarenal complications of the nephrotic syndrome. *Am J Kidney Dis* 23: 477–497, 1994.
- 62. Hazell LJ, and Stocker R. α -Tocopherol does not inhibit hypochlorite-induced oxidation of apolipoprotein B-100 of low-density lipoprotein. *FEBS Lett* 414: 541–544, 1997.
- 63. Heinecke JW, Hsu FF, Crowley JR, Hazen SL, Leeuwenburgh C, Mueller DM, Rasmussen JE, and Turk J. Detecting oxidative modification of biomolecules with isotope dilution mass spectrometry: sensitive and quantitative assays for oxidized amino acids in proteins, and tissues. *Methods Enzymol* 300: 124–144, 1998.
- 64. Horstmann HJ, Rohen JW, and Sames K. Age-related changes in the composition of proteins in the trabecular mesh-work of the human eye. *Mech Ageing Dev* 21: 121–136, 19803.
- 65. Irwin JA, Ostdal H, and Davies MJ. Myoglobin-induced oxidative damage: evidence for radical transfer from oxidized myoglobin to other proteins and antioxidants. *Arch Biochem Biophys* 362: 94–104, 1999.
- 66. Lazo JS, and Pitt BR. Metallothioneins and cell death by anticancer drugs. *Annu Rev Pharmacol Toxicol* 35: 635–653, 1995.
- 67. Lazo JS, Kondo Y, Dellapiazza D, Michalska AE, Choo KH, and Pitt BR. Enhanced sensitivity to oxidative stress in cultured embryonic cells from transgenic mice deficient in metallothionein I and II genes. *J Biol Chem* 270: 5506–5510, 1995.
- 68. Leeuwenburgh C, Hardy MM, Hazen SL, Wagner P, Oh-ishi S, Steinbrecher UP, and Heinecke JW. Reactive nitrogen intermediates promote low density lipoprotein oxidation in human atherosclerotic intima. *J Biol Chem* 272: 1433–1436, 1997.
- 69. Leeuwenburgh C, Hansen PA, Holloszy JO, and Heinecke JW. Oxidized amino acids in the urine of aging rats: potential markers for assessing oxidative stress in vivo. *Am J Physiol* 276: R128–R135, 1999.
- Levine RL, Mosoni L, Berlett BS, and Stadtman ER. Methionine residues as endogenous antioxidants in proteins. *Proc Natl Acad Sci U S A* 93: 15036–15040, 1996.
- 71. Lewisch SA, and Levine RL. Determination of 2–oxohistidine by amino acid analysis. *Methods Enzymol* 300: 120–124, 1999.
- 72. Luoma J, Näyhä S, Sikkilä K, and Hassi J. High serum alpha-tocopherol, albumin, selenium and

- cholesterol, and mortality from coronary heart disease in northern Finland. *J Intern Med* 237: 49–54, 1995.
- 73. Mander EL, Dean RT, Stanley KK, and Jessup W. Apolipoprotein B of oxidized LDL accumulates in the lysosomes of macrophages. *Biochim Biophys Acta* 1212: 80–92, 1994.
- 74. McCully KS. Homocysteine theory of arteriosclerosis development and current status. *Atheroscler Rev* 11: 157–246, 1983.
- 75. Miller NJ, Rice-Evans C, Davies MJ, Gopinathan V, and Milner A. A novel method for measuring antioxidant capacity and its application to monitoring the antioxidant status in premature neonates. *Clin Sci* 84: 407–412, 1993.
- Moller P, Wallin H, and Knudsen LE. Oxidative stress associated with exercise, psychological stress and life-style factors. *Chem Biol Interact* 102: 17–36, 1996.
- 77. Montgomery DAD, Neill DW, and Dowdle EBD. Idiopathic hypoalbuminemia. *Clin Sci* 22: 141–154, 1962.
- 78. Moskovitz J, Weissbach H, and Brot N. Cloning the expression of a mammalian gene involved in the reduction of methionine sulfoxide residues in proteins. *Proc Natl Acad Sci U S A* 93: 2095–2099, 1996.
- 79. Moskovitz J, Flescher E, Berlett BS, Azare J, Poston JM, and Stadtman ER. Overexpression of peptidemethionine sulfoxide reductase in *Saccharomyces cerevisiae* and human T cell provides them with high resistance to oxidative stress. *Proc Natl Acad Sci U S A* 95: 14071–14075, 1997.
- 80. Navab M, Berliner JA, Watson AD, Hama SY, Territo MC, Lusis AJ, Shih DM, Van Lenten BJ, Frank JS, Demer LL, Edwards PA, and Fogelman AM. The yin and yang of oxidation in the development of the fatty streak. *Arterioscler Thromb Vasc Biol* 16: 831–842, 1996
- 81. Ogino T, and Okada S. Oxidative damage of bovine serum albumin and other enzyme proteins by iron-chelate complexes. *Biochim Biophys Acta* 1245: 359–365, 1995.
- 82. Olszewski AJ, and McCully KS. Homocysteine metabolism and the oxidative modification of proteins and lipids. *Free Radic Biol Med* 14: 683–693, 1993.
- 83. Pacifici RE, and Davies KJA. Protein, lipid and DNA repair systems in oxidative stress: The free-radical theory of aging revisited. *Gerontology* 37: 166–180, 1991
- 84. Peters T Jr. *All about Albumin: Biochemistry, Genetics, and Medical Applications*. London, U.K.: Academic Press Ltd., 1996.
- 85. Phillips A, Shaper AG, and Whincup PH. Association between serum albumin and mortality from cardiovascular disease, cancer and other causes. *Lancet* 16: 1434–1436, 1989.
- 86. Pitt BR, Schwarz M, Woo ES, Yee E, Wasserloos K, Tran S, Weng W, Mannix RJ, Watkins SA, Tyurina YY, Tyurin VA, Kagan VE, and Lazo JS. Overexpression of metallothionein decreases sensitivity of

- pulmonary endothelial cells to oxidant injury. *Am J Physiol* 273: L856–L865, 1997.
- 87. Prost M. Process for the determination by means of free radicals of the antioxidant properties of a living organism or a potentially aggressive agents. U.S. Patent No. 576,460(5,135,850). 1992. 4-8-1992.
- 88. Pryor WA, and Stone K. Oxidants in cigarette smoke: radicals, hydrogen peroxide, peroxynitrate, and peroxynitrite. *Ann N Y Acad Sci* 686: 12–28, 1993.
- 89. Quesada AR, Byrnes RW, Krezoski SO, and Petering DH. Direct reaction of H₂O₂ with sulfhydryl groups in HL-60 cells: zinc-metallothionein and other sites. *Arch Biochem Biophys* 334: 241–250, 1996.
- Quinlan GJ, Margarson MP, Mumby S, Evans TW, and Gutteridge JMC. Administration of albumin to patients with sepsis syndrome: a possible beneficial role in plasma thiol repletion. Clin Sci 95: 459–465, 1998.
- 91. Rahman MA, Nelson H, Weissbach H, and Brot N. Cloning, sequencing, and expression of the *Escherichia coli* peptide methionine sulfoxide reductase gene. *J Biol Chem* 267: 15549–15551, 1992.
- 92. Rajeswari P, Natarajan R, Nadler JL, Kumar D, and Kalra VK. Glucose induces lipid peroxidation and inactivation of membrane-associated ion-transport enzymes in human erythrocytes in vivo and in vitro. *J Cell Physiol* 149: 100–109, 1991.
- 93. Ravichandran V, Seres T, Moriguchi T, Thomas JA, and Johnson RB. *S*-Thiolation of glyceraldehyde-3–phosphate dehydrogenase induced by phagocytosis-associated respiratory burst in blood monocytes. *J Biol Chem* 269: 25010–25015, 1994.
- Reddy VY, Desrochers PE, Pizzo SV, Gonias SL, Sahakian JA, Levine RL, and Weiss SJ. Oxidative dissociation of human α2-macroglobulin tetramers into dysfunctional dimers. *J Biol Chem* 269: 4683–4691, 1994
- 95. Rivett AJ. The multicatalytic proteinase of mammalian cells. *Arch Biochem Biophys* 268: 1–8, 1989.
- 96. Rokutan K, Thomas JA, and Johnston RB Jr. Phagocytosis and stimulation of the respiratory burst by phorbol diester initiate *S*-thiolation of specific proteins in macrophages. *J Immunol* 147: 260–264, 1991.
- 97. Rosenblatt DS: Inherited disorders of folate transport and metabolism. In: *Metabolic Basis of Inherited Disease*, edited by Scriver CR, Beaudet AL, Sly WS, and Valle D. McGraw-Hill, New York: 1995, pp. 3111–3128.
- 98. Rothblat GH, Mahlberg FH, Johnson WJ, and Phillips MC. Apolipoproteins, membrane cholesterol domains, and the regulation of cholesterol efflux. *J Lipid Res* 33: 1091–1097, 1992.
- 99. Sadanandom A, Piffanelli P, Knott T, Robinson C, Sharpe A, Lydiate D, Murphy D, and Fairbairn DJ. Identification of a peptide methionine sulphoxide reductase gene in an oleosin promoter from *Brassica napus*. *Plant J* 10: 235–242, 1996.
- 100. Saintot M, Bernard N, Astre C, Gerber M, and Cerber M. Ozone exposure and blood antioxidants: A study in a periurban area in Southern France. *Arch Environ Health* 54: 34–39, 1999.

- 101. Salo DC, Pacifici RE, Lin SW, Giulivi C, and Davies KJA. Superoxide dismutase undergoes proteolysis and fragmentation following oxidative modification and inactivation. J Biol Chem 265: 11919–11927, 1990.
- Sato M, and Bremner I. Oxygen free radicals and metallothionein. Free Radic Biol Med 14: 325–337, 1993.
- 103. Schwarz MA, Lazo JS, Yalowich JC, Reynolds I, Kagan VE, Tyurin V, Kim YM, Watkins SC, and Pitt BR. Cytoplasmic metallothionein overexpression protects NIH 3T3 cells from *tert*-butyl hydroperoxide toxicity. *J Biol Chem* 269: 15238–15243, 1994.
- 104. Schwarz MA, Lazo JS, Yalowich JC, Allen WP, Whitmore M, Bergonia HA, Tzeng E, Billiar TR, Robbins PD, and Lancaster JR Jr. Metallothionein protects against the cytotoxic and DNA-damaging effects of nitric oxide. *Proc Natl Acad Sci U S A* 92: 4452–4456, 1995.
- 105. Shacter E, Williams JA, and Levine RL. Oxidative modification of fibrinogen inhibits thrombin-catalyzed clot formation. Free Radic Biol Med 18: 815–821, 1995.
- 106. Shaklai N, Garlick RL, and Bunn HF. Nonenzymic glycosylation of human serum albumin alters its conformation and function. *J Biol Chem* 259: 3812–3817, 1984.
- 107. Smith WL, Eling TE, Kulmacz RJ, Marnett LJ, and Tsai A. Tyrosyl radicals and their role in hydroper-oxide-dependent activation and inactivation of prostaglandin endoperoxide synthase. *Biochemistry* 31: 3–7, 1992.
- 108. Soszynski M, and Bartosz G. Decrease in accessible thiols as an index of oxidative damage to membrane proteins. *Free Radic Biol Med* 23: 463–469, 1997.
- 109. Stadtman ER. Oxidation of free amino acids and amino acid residues in proteins by radiolysis and by metal-catalyzed reactions. *Ann Rev Biochem* 62: 797–821, 1993.
- 110. Stafforini DM, Zimmerman GA, McIntyre TM, and Prescott SM. The plasma PAF acetylhydrolase prevents oxidative modification of low density lipoprotein. *J Lipid Mediat* 10: 53–56, 1994.
- 111. Steinberg D, and Lewis A. Conner Memorial Lecture—Oxidative modification of LDL and atherogenesis. *Circulation* 95: 1062–1071, 1997.
- 112. Stocchi V, Biagiarelli B, Fiorani M, Palma F, Piccoli G, Cucchiarini L, and Dachà M. Inactivation of rabbit red blood cell hexokinase activity promoted *in vitro* by an oxygen-radical-generating system. *Arch Biochem Biophys* 311: 160–167, 1994.
- 113. Templeton DM, and Cherian MG. Toxicological significance of metallothionein. *Methods Enzymol* 205: 11–24, 1991.
- 114. Terada LS, Leff JA, Guidot DM, Willingham IR, and Repine JE. Inactivation of xanthine oxidase by hydrogen peroxide involves site-directed hydroxyl radical formation. *Free Radic Biol Med* 10: 61–68, 1991.
- 115. Thomas JA, Poland B, and Honzatko R. Protein sulfhydryls and their role in the antioxidant function of protein *S*-thiolation. *Arch Biochem Biophys* 319: 1–9, 1995.

- 116. Uchida K, Kanematsu M, Sakai K, Matsuda T, Hattori N, Mizuno Y, Suzuki D, Miyata T, Noguchi N, Niki E, and Osawa T. Protein-bound acrolein: potential markers for oxidative stress. *Proc Natl Acad Sci U S A* 95: 4882–4887, 1998.
- 117. Vander Jagt DL, Hunsaker LA, Vander Jagt TJ, Gomez MS, Gonzales DM, Deck LM, and Royer RE. Inactivation of glutathione reductase by 4–hydroxynonenal and other endogenous aldehydes. *Biochem Pharmacol* 53: 1133–1140, 1997.
- 118. Van Kuijk FJGM, Sevanian A, Handelman GJ, and Dratz EA. A new role for phospholipase A2: protection of membrane from lipid peroxidation damage. *Trends Biochem Sci* 12: 31–34, 1987.
- 119. Vasak M, and Hasler DW. Metallothioneins: new functional and structural insights. *Curr Opin Chem Biol* 4: 177–183, 2000.
- 120. Vlassara H. Advanced glycation end-products and atherosclerosis. *Ann Med* 28: 419–426, 1996.
- 121. Vlassara H, Brownlee M, and Cerami A. High-affinity receptor-mediated uptake and degradation of glucose-modified proteins: a potential mechanism for the removal of senescent macromolecules. *Proc Natl Acad Sci U S A* 82: 5588–5592, 1985.
- 122. Vogt W. Oxidation of methionyl residues in proteins: tools, targets, and reversal. *Free Radic Biol Med* 18: 93–105, 1995.
- 123. Watson AD, Navab M, Hama SY, Sevanian A, Prescott SM, Stafforini DM, McIntyre TM, La Du BN, Fogelman AM, and Berliner JA. Effect of platelet activating factor-acetylhydrolase on the formation and action of minimally oxidized low density lipoprotein. *J Clin Invest* 95: 774–782, 1995.
- 124. Wayner DDM, Burton GW, Ingold KU, and Locke SJ. Quantitative measurement of total, peroxyl radi-

- cal-trapping antioxidant capability of human blood plasma by controlled peroxidation. *FEBS Lett* 187: 33–37, 1985.
- 125. Wheeler DC, and Bernard DB. Lipid abnormalities in the nephrotic syndrome: causes, consequences, and treatment. *Am J Kidney Dis* 23: 331–346, 1994.
- 126. Wolff SP, Jiang ZY, and Hunt JV. Protein glycation and oxidative stress in diabetes mellitus and ageing. *Free Radic Biol Med* 10: 339–352, 1991.
- 127. Woodside JV, Yarnell JWG, McMaster D, Young IS, Harmon DL, McCrum EE, Patterson CC, Gey KF, Whitehead AS, and Evans A. Effect of B-group vitamins and antioxidant vitamins on hyperhomocysteinemia: a double-blind, randomized, factorial-design, controlled trial. *Am J Clin Nutr* 67: 858–866, 1998.
- 128. Zou M, Jendral M, and Ullrich V. Prostaglandin endoperoxide-dependent vasospasm in bovine coronary arteries after nitration of prostacyclin synthase. *Br J Pharmacol* 126: 1283–1292, 1999.

Address reprint requests to:
Dr. Denis Blache
INSERM-INRA
Unité de Nutrition Lipidique
17, rue Sully BP 86510
21065 Dijon, France

E-mail: blache@dijon.inra.fr

Received for publication March 24, 2000; accepted November 10, 2000.

This article has been cited by:

- 1. Hasmik Grigoryan, He Li, Anthony T. Iavarone, Evan R. Williams, Stephen M. Rappaport. 2012. Cys34 Adducts of Reactive Oxygen Species in Human Serum Albumin. *Chemical Research in Toxicology* **25**:8, 1633-1642. [CrossRef]
- 2. Gabriella Fanali, Alessandra di Masi, Viviana Trezza, Maria Marino, Mauro Fasano, Paolo Ascenzi. 2012. Human serum albumin: From bench to bedside. *Molecular Aspects of Medicine* **33**:3, 209-290. [CrossRef]
- 3. Sema Demirci Çekiç, Nilay Kara, Esma Tütem, Kevser Sözgen Ba#kan, Re#at Apak. 2012. Protein–Incorporated Serum Total Antioxidant Capacity Measurement by a Modified CUPRAC (CUPRIC Reducing Antioxidant Capacity) Method. *Analytical Letters* **45**:7, 754-763. [CrossRef]
- 4. Magdalene K. Montgomery, William A. Buttemer, A.J. Hulbert. 2012. Does the oxidative stress theory of aging explain longevity differences in birds? II. Antioxidant systems and oxidative damage. *Experimental Gerontology*. [CrossRef]
- 5. Yasunori Iwao, Yu Ishima, Junji Yamada, Taishi Noguchi, Ulrich Kragh-Hansen, Katsumi Mera, Daisuke Honda, Ayaka Suenaga, Toru Maruyama, Masaki Otagiri. 2012. Quantitative evaluation of the role of cysteine and methionine residues in the antioxidant activity of human serum albumin using recombinant mutants. *IUBMB Life* n/a-n/a. [CrossRef]
- 6. Kwang Joon Kim, Byung-Wan Lee. 2012. The Roles of Glycated Albumin as Intermediate Glycation Index and Pathogenic Protein. *Diabetes & Metabolism Journal* **36**:2, 98. [CrossRef]
- 7. Akhlaq A. FarooquiGeneration of Reactive Oxygen Species in the Brain: Signaling for Neural Cell Survival or Suicide 1-15. [CrossRef]
- 8. ALAN D. PEMBERTON, JEREMY K. BROWN, NICKY M. CRAIG, JUDITH PATE, KEVIN McLEAN, NEIL F. INGLIS, DAVID KNOX, PAMELA A. KNIGHT. 2011. Changes in protein expression in the sheep abomasum following trickle infection with Teladorsagia circumcincta. *Parasitology* 1-11. [CrossRef]
- 9. Valerio Chiurchiù, Mauro Maccarrone. 2011. Chronic Inflammatory Disorders and Their Redox Control: From Molecular Mechanisms to Therapeutic Opportunities. *Antioxidants & Redox Signaling* 15:9, 2605-2641. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF] with Links]
- 10. Su Yin Lim, Mark J. Raftery, Carolyn L. Geczy. 2011. Oxidative Modifications of DAMPs Suppress Inflammation: The Case for S100A8 and S100A9. *Antioxidants & Redox Signaling* **15**:8, 2235-2248. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF with Links]
- 11. László Selmeci. 2011. Advanced oxidation protein products (AOPP): novel uremic toxins, or components of the non-enzymatic antioxidant system of the plasma proteome?. *Free Radical Research* **45**:10, 1115-1123. [CrossRef]
- 12. Tiago R. Figueira, Roger F. Castilho, Ângela Saito, Helena C.F. Oliveira, Anibal E. Vercesi. 2011. The higher susceptibility of congenital analbuminemic rats to Ca2+-induced mitochondrial permeability transition is associated with the increased expression of cyclophilin D and nitrosothiol depletion. *Molecular Genetics and Metabolism*. [CrossRef]
- 13. Letícia Filippon, Camila S. Vanzin, Giovana B. Biancini, Izabela N. Pereira, Vanusa Manfredini, Angela Sitta, Maria do Carmo R. Peralba, Ida V.D. Schwartz, Roberto Giugliani, Carmen R. Vargas. 2011. Oxidative stress in patients with mucopolysaccharidosis type II before and during enzyme replacement therapy. *Molecular Genetics and Metabolism* 103:2, 121-127. [CrossRef]
- 14. Tomoe Kugimiya, Hirofumi Jono, Shiori Saito, Toru Maruyama, Daisuke Kadowaki, Yohei Misumi, Yoshinobu Hoshii, Masayoshi Tasaki, Yu Su, Mitsuharu Ueda, Konen Obayashi, Makoto Shono, Masaki Otagiri, Yukio Ando. 2011. Loss of functional albumin triggers acceleration of transthyretin amyloid fibril formation in familial amyloidotic polyneuropathy. *Laboratory Investigation*. [CrossRef]
- 15. Philippe Rondeau, Emmanuel Bourdon. 2011. The glycation of albumin: Structural and functional impacts. *Biochimie* **93**:4, 645-658. [CrossRef]

- 16. Anna Maria Salzano, Giovanni Renzone, Andrea Scaloni, Armida Torreggiani, Carla Ferreri, Chryssostomos Chatgilialoglu. 2011. Human serum albumin modifications associated with reductive radical stress. *Molecular BioSystems* 7:3, 889. [CrossRef]
- 17. Franziska Bosshard, Kathrin Riedel, Thomas Schneider, Carina Geiser, Margarete Bucheli, Thomas Egli. 2010. Protein oxidation and aggregation in UVA-irradiated Escherichia coli cells as signs of accelerated cellular senescence. *Environmental Microbiology* **12**:11, 2931-2945. [CrossRef]
- 18. Jelena M. A#imovi#, Bojana D. Stanimirovi#, Nina Todorovi#, Vesna B. Jovanovi#, Ljuba M. Mandi#. 2010. Influence of the microenvironment of thiol groups in low molecular mass thiols and serum albumin on the reaction with methylglyoxal. *Chemico-Biological Interactions* **188**:1, 21-30. [CrossRef]
- 19. Hiroyuki Terawaki, Yukie Takada, Seiichi Era, Yoichi Funakoshi, Keisuke Nakayama, Masaaki Nakayama, Makoto Ogura, Sadayoshi Ito, Tatsuo Hosoya. 2010. The Redox State of Albumin and Serious Cardiovascular Incidence in Hemodialysis Patients. *Therapeutic Apheresis and Dialysis* 14:5, 465-471. [CrossRef]
- 20. F. Bosshard, M. Bucheli, Y. Meur, T. Egli. 2010. The respiratory chain is the cell's Achilles' heel during UVA inactivation in Escherichia coli. *Microbiology* **156**:7, 2006-2015. [CrossRef]
- 21. Graziano Colombo, Giancarlo Aldini, Marica Orioli, Daniela Giustarini, Rosalba Gornati, Ranieri Rossi, Roberto Colombo, Marina Carini, Aldo Milzani, Isabella Dalle-Donne. 2010. Water-Soluble #,#-Unsaturated Aldehydes of Cigarette Smoke Induce Carbonylation of Human Serum Albumin. *Antioxidants & Redox Signaling* 12:3, 349-364. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF with Links]
- 22. Endry Nugroho Prasetyo, Tukayi Kudanga, Walter Steiner, Michael Murkovic, Gibson S. Nyanhongo, Georg M. Guebitz. 2010. Laccase-generated tetramethoxy azobismethylene quinone (TMAMQ) as a tool for antioxidant activity measurement. *Food Chemistry* **118**:2, 437-444. [CrossRef]
- 23. Sushma K Cribbs, Greg S Martin. 2009. Fluid balance and colloid osmotic pressure in acute respiratory failure: optimizing therapy. *Expert Review of Respiratory Medicine* **3**:6, 651-662. [CrossRef]
- 24. T. Cindrova-Davies. 2009. Gabor Than Award Lecture 2008: Pre-eclampsia From Placental Oxidative Stress to Maternal Endothelial Dysfunction. *Placenta* **30**, 55-65. [CrossRef]
- 25. Peter Rapta, Katarína Valachová, Peter Gemeiner, Ladislav S#oltés. 2009. High-Molar-Mass Hyaluronan Behavior During Testing Its Radical Scavenging Capacity in Organic and Aqueous Media: Effects of the Presence of Manganese(II) Ions. *Chemistry & Biodiversity* **6**:2, 162-169. [CrossRef]
- 26. Philippe Rondeau, Nihar Ranjan Singh, Henri Caillens, Frank Tallet, Emmanuel Bourdon. 2008. Oxidative stresses induced by glycoxidized human or bovine serum albumin on human monocytes. *Free Radical Biology and Medicine* **45**:6, 799-812. [CrossRef]
- 27. Dunyaporn Trachootham , Weiqin Lu , Marcia A. Ogasawara , Nilsa Rivera-Del Valle , Peng Huang . 2008. Redox Regulation of Cell Survival. *Antioxidants & Redox Signaling* **10**:8, 1343-1374. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF with Links]
- 28. A. Marin, G. Pozza, F. Gottardo, L. Moro, A. L. Stefani, G. Cozzi, M. Brscic, I. Andrighetto, L. Ravarotto. 2008. Administration of dexamethasone per os in finishing bulls. II. Effects on blood parameters used as indicators of animal welfare. *animal* 2:07. . [CrossRef]
- 29. Marjolaine Roche, Philippe Rondeau, Nihar Ranjan Singh, Evelyne Tarnus, Emmanuel Bourdon. 2008. The antioxidant properties of serum albumin. *FEBS Letters* **582**:13, 1783-1787. [CrossRef]
- 30. H. M. Erdogan, M. Karapehlivan, M. Citil, O. Atakisi, E. Uzlu, A. Unver. 2008. Serum sialic acid and oxidative stress parameters changes in cattle with leptospirosis. *Veterinary Research Communications* 32:4, 333-339. [CrossRef]
- 31. Toshio Okazaki, Noriyuki Okudaira, Naohito Ishii, Hiroshi Yotsuyanagi, Toshiaki Nagai, Shougo Tokudome, Takahiro Fujioka, Shinichiro Takahashi. 2008. Comparison of the Antioxidant Activity of Albumin from Various Animal Species. *Zoological Science* 25:2, 172-177. [CrossRef]
- 32. Philippe Rondeau, Sergio Armenta, Henri Caillens, Serge Chesne, Emmanuel Bourdon. 2007. Assessment of temperature effects on #-aggregation of native and glycated albumin by FTIR

- spectroscopy and PAGE: Relations between structural changes and antioxidant properties. *Archives of Biochemistry and Biophysics* **460**:1, 141-150. [CrossRef]
- 33. Dr. Denis Blache, Suzanne Lussier–Cacan, Jacques Gagnon, Arthur S. Leon, D.C. Rao, James S. Skinner, Jack H. Wilmore, Tuomo Rankinen, Claude Bouchard, Jean Davignon. 2007. Effect of Exercise Training on In Vitro LDL Oxidation and Free Radical–Induced Hemolysis: The HERITAGE Family Study. *Antioxidants & Redox Signaling* 9:1, 123-130. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 34. M ALMAJANO, M DELGADO, M GORDON. 2007. Changes in the antioxidant properties of protein solutions in the presence of epigallocatechin gallate. *Food Chemistry* **101**:1, 126-130. [CrossRef]
- 35. Serge Chesne, Philippe Rondeau, Sergio Armenta, Emmanuel Bourdon. 2006. Effects of oxidative modifications induced by the glycation of bovine serum albumin on its structure and on cultured adipose cells. *Biochimie* **88**:10, 1467-1477. [CrossRef]
- 36. Denis Blache, Sylvie Devaux, Olivier Joubert, Nadine Loreau, Martina Schneider, Philippe Durand, Michel Prost, Vincent Gaume, Markus Adrian, Pascal Laurant, Alain Berthelot. 2006. Long-term moderate magnesium-deficient diet shows relationships between blood pressure, inflammation and oxidant stress defense in aging rats. *Free Radical Biology and Medicine* **41**:2, 277-284. [CrossRef]
- 37. Bruce Ames, Jiankang LiuMitochondrial Nutrients 20052039, 59-105. [CrossRef]
- 38. M-H Tsai, Y-C Chen, C-S Wu, Y-P Ho, J-T Fang, J-M Lien, C. Yang, Y-Y Chu, N-J Liu, C-H Lin, C-T Chiu, P-C Chen. 2005. Extracorporal liver support with molecular adsorbents recirculating system in patients with hepatitis B-associated fulminant hepatic failure. *International Journal of Clinical Practice* **59**:11, 1289-1294. [CrossRef]
- 39. Karol A. Mathews, Maureen Barry. 2005. The use of 25% human serum albumin: outcome and efficacy in raising serum albumin and systemic blood pressure in critically ill dogs and cats. *Journal of Veterinary Emergency and Critical Care* **15**:2, 110-118. [CrossRef]
- 40. 2003. Trend of Most Cited Papers (2001-2002) in ARS. *Antioxidants & Redox Signaling* **5**:6, 813-815. [Citation] [Full Text PDF] [Full Text PDF with Links]
- 41. Kinga A. Powers, Andras Kapus, Rachel G. Khadaroo, Ruijuan He, John C. Marshall, Thomas F. Lindsay, Ori D. Rotstein. 2003. Twenty-five percent albumin prevents lung injury following shock/resuscitation. *Critical Care Medicine* 31:9, 2355-2363. [CrossRef]
- 42. Mohammed Habdous, Bernard Herbeth, Monique Vincent-Viry, John V. Lamont, Peters S. Fitzgerald, Sophie Visvikis, Gérard Siest. 2003. Serum Total Antioxidant Status, Erythrocyte Superoxide Dismutase and Whole-Blood Glutathione Peroxidase Activities in the Stanislas Cohort: Influencing Factors and Reference Intervals. *Clinical Chemistry and Laboratory Medicine* 41:2, 209-215. [CrossRef]
- 43. Takashi ASHINO, Shigenari OZAWA, Satoshi NUMAZAWA, Takemi YOSHIDA. 2003. TISSUE-DEPENDENT INDUCTION OF HEME OXYGENASE-1 AND METALLOTHIONEIN-1/2 BY METHYL METHANESULFONATE. *The Journal of Toxicological Sciences* 28:3, 181-189. [CrossRef]
- 44. Rajiv Jalan, Sambit Sen, Christian Steiner, Dharmesh Kapoor, Akeel Alisa, Roger Williams. 2003. Extracorporeal liver support with molecular adsorbents recirculating system in patients with severe acute alcoholic hepatitis. *Journal of Hepatology* **38**:1, 24-31. [CrossRef]
- 45. Chandan K. Jana, Nilanjana Das, Rajindar S. Sohal. 2002. Specificity of Age-Related Carbonylation of Plasma Proteins in the Mouse and Rat. *Archives of Biochemistry and Biophysics* **397**:2, 433-439. [CrossRef]